

101.126-12 – including *Taq* polymerase, IFU-01  
101.126-12u – without *Taq* polymerase, IFU-02

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“Instructions for Use” (IFU)

Lot No.: **60X**

Lot-specific information  
**Olerup SSP® DRB1\*16**

Product number:	101.126-12 – including <i>Taq</i> polymerase 101.126-12u – without <i>Taq</i> polymerase
Lot number:	60X
Expiry date:	2017-May-01
Number of tests:	12
Number of wells per test:	15+1
Storage - pre-aliquoted primers:	dark at -20°C
- PCR Master Mix:	-20°C
- Adhesive PCR seals	RT
- Product Insert	RT

**This Product Description is only valid for Lot No. 60X.**

Complete product documentation consists of generic Instructions for Use (IFU), lot specific Product Insert, Worksheet and Certificate.

**CHANGES COMPARED TO THE PREVIOUS OLERUP SSP®  
DRB1\*16 LOT (84R)**

The DRB1\*16 kit is updated for new alleles to enable separation of:

- Confirmed<sup>1</sup> alleles as listed in the IMGT/HLA database
- Polymorphisms in exons outside of the region encoding the peptide binding domain
- Null and Alternatively expressed alleles

A well containing Negative Control primer pairs has been added.

The format of the Product Insert and Worksheet have been changed.

<sup>1</sup>As described in section Uniquely Identified Alleles.

The DRB1\*16 primer set, specificity and interpretation tables have been updated for the DRB1 alleles described since the previous *Olerup SSP®* DRB1\*16 lot was made (**Lot No. 84R**). The kit design is based on IMGT/HLA database 3.17.0.

The primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

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As of lot series V, the Specificity Table is included in the lot-specific Product Insert, and the Interpretation Table is included in the Worksheet.

The primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

Well	5'-primer	3'-primer	rationale
8	-	Added	3'-primer added from well 14.
11	Added	-	5'-primer added for the DRB1*16:21N allele.
13	-	Added	3'-primer added from well 14.
14	Added	Added	Primer pairs added from well 16.
16	Moved	Moved	Primer pairs moved to well 14, negative control.

Change in revision R01 compared to R00:

1. The DRB1\*16:12 allele is amplified by primer mixes 1 and 2, and is weakly amplified by primer mix 12. The DRB1\*15:28 allele is amplified by primer mix 9. This has been corrected in the Specificity and Interpretation Tables.

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Well **16** contains Negative Control primer pairs, that will amplify more than 95% of the *Olerup SSP®* HLA Class I, DRB, DQB1, DPB1 and DQA1 amplicons as well as all the amplicons generated by the control primer pairs matching the human growth hormone gene.

HLA-specific PCR product sizes range from 75 to 200 base pairs.  
The PCR product generated by the positive control primer pair is 430 base pairs.

Length of PCR product	105	200	105	80	75	80	85
<b>5'-primer<sup>1</sup></b>	<b>164</b>	<b>340</b>	<b>440</b>	<b>45</b>	<b>45</b>	<b>43</b>	<b>36</b>
	5'-CAC <sup>3'</sup>	5'-Agg <sup>3'</sup>	5'-TTA <sup>3'</sup>	5'-Tgg <sup>3'</sup>	5'-Tgg <sup>3'</sup>	5'-Tgg <sup>3'</sup>	5'-TAC <sup>3'</sup>
							<b>36</b>
							5'-TAT <sup>3'</sup>
<b>3'-primer<sup>2</sup></b>	<b>231</b>	<b>2<sup>nd</sup> I</b>	<b>507</b>	<b>59</b>	<b>58</b>	<b>57</b>	<b>47</b>
	5'-TgC <sup>3'</sup>	5'-AAA <sup>3'</sup>	5'-TTg <sup>3'</sup>	5'-CTC <sup>3'</sup>	5'-ggC <sup>3'</sup>	5'-CTC <sup>3'</sup>	5'-ACA <sup>3'</sup>
							<b>48</b>
							5'-gCA <sup>3'</sup>
							<b>48</b>
							5'-gCC <sup>3'</sup>
							<b>52</b>
							5'-TgT <sup>3'</sup>
<b>A*</b>	+	+	+				
<b>B*</b>	+	+	+				
<b>C*</b>	+	+	+				
<b>DRB1</b>				+	+		
<b>DRB3</b>				+	+		
<b>DRB5</b>				+			
<b>DQB1</b>					+		
<b>DPB1</b>						+	
<b>DQA1</b>							+

<sup>1</sup>The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2<sup>nd</sup> or 3<sup>rd</sup> exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

<sup>2</sup>The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2<sup>nd</sup> or 3<sup>rd</sup> exon or the 2<sup>nd</sup> intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

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## PRODUCT DESCRIPTION

### DRB1\*16 SSP subtyping

#### CONTENT

The primer set contains 5'- and 3'-primers for identifying the DRB1\*16:01 to DRB1\*16:24 alleles.

#### PLATE LAYOUT

Each test consists of 16 PCR reactions in a 16 well cut PCR plate.

<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>	<b>6</b>	<b>7</b>	<b>8</b>
<b>9</b>	<b>10</b>	<b>11</b>	<b>12</b>	<b>13</b>	<b>14</b>	<b>15</b>	<b>NC</b>

The 16 well cut PCR plate is marked with 'DRB1\*16' in silver/gray ink.

Well No. 1 is marked with the Lot No. '60X'.

Wells 1 to 15 – DRB1\*16 high resolution primers.

Well 16 – Negative Control (NC).

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded.

The PCR plates are covered with a PCR-compatible foil.

**Please note:** When removing each 16 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

#### INTERPRETATION

Due to the sharing of sequence motifs between DRB1 alleles, primer mixes 1, 5, 6, 9, 10, 14 and 15 will amplify other DRB1 alleles. In addition, DRB5 alleles will be amplified by primer mix 14.

For further details see Specificity Table.

#### UNIQUELY IDENTIFIED ALLELES

All the DRB1\*16 alleles, i.e. **DRB1\*16:01 to DRB1\*16:24**, recognized by the HLA Nomenclature Committee in July 2014<sup>1,2</sup> will be amplified by the primers in the DRB1\*16 subtyping kit.

The DRB1\*16 kit enables separation of the confirmed DRB1\*16 alleles as listed in the IMGT/HLA database. An HLA allele is listed as confirmed by IMGT/HLA if it has been sequenced by more than a single laboratory or from multiple sources. Current allele confirmation status for DRB1\*16 alleles is listed below.

The DRB1\*16 kit also enables identification of polymorphisms in exons outside of the region encoding the peptide binding domain and of null and alternatively expressed alleles.

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The DRB1\*16 subtyping kit cannot distinguish the following silent mutations: the DRB1\*16:01:01-16:01:05, DRB1\*16:02:01-16:02:03, the DRB1\*16:05:01-16:05:02 or the DRB1\*16:09:01-16:09:02 alleles.

<sup>1</sup>DRB1 alleles listed on the IMGT/HLA web page 2014-July-25, release 3.17.0, [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla).

<sup>2</sup>Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page <http://hla.alleles.org/alleles/deleted.html>.

## ALLELE CONFIRMATION STATUS

Allele	Status <sup>1</sup>	Allele	Status <sup>1</sup>
<b>DRB1*16:01:01</b>	<b>Confirmed</b>	<b>DRB1*16:14</b>	<b>Confirmed</b>
DRB1*16:01:02	Unconfirmed	<b>DRB1*16:15</b>	<b>Confirmed</b>
DRB1*16:01:03	Unconfirmed	DRB1*16:16	Unconfirmed
DRB1*16:01:04	Unconfirmed	DRB1*16:17	Unconfirmed
<b>DRB1*16:01:05</b>	<b>Confirmed</b>	DRB1*16:18	Unconfirmed
<b>DRB1*16:02:01</b>	<b>Confirmed</b>	DRB1*16:19	Unconfirmed
DRB1*16:02:02	Unconfirmed	DRB1*16:20	Unconfirmed
DRB1*16:02:03	Unconfirmed	DRB1*16:21N	Unconfirmed
DRB1*16:03	Unconfirmed	DRB1*16:22	Unconfirmed
<b>DRB1*16:04</b>	<b>Confirmed</b>	DRB1*16:23	Unconfirmed
<b>DRB1*16:05:01</b>	<b>Confirmed</b>	DRB1*16:24	Unconfirmed
<b>DRB1*16:05:02</b>	<b>Confirmed</b>		
DRB1*16:07	Unconfirmed		
<b>DRB1*16:08</b>	<b>Confirmed</b>		
<b>DRB1*16:09:01</b>	<b>Confirmed</b>		
DRB1*16:09:02	Unconfirmed		
<b>DRB1*16:10</b>	<b>Confirmed</b>		
DRB1*16:11	Unconfirmed		
<b>DRB1*16:12</b>	<b>Confirmed</b>		
DRB1*16:13N	Unconfirmed		

<sup>1</sup>Allele status “confirmed” or “unconfirmed” as listed on the IMGT/HLA web page 2014-July-25, release 3.17.0, [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla).

## RESOLUTION IN HOMO- AND HETEROZYGOTES

Results file with resolution in DRB1\*16 homo- and heterozygotes is available upon request.

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Lot-specific information  
**SPECIFICITY TABLE**

**DRB1\*16 SSP subtyping**

**Specificities and sizes of the PCR products of the 15+1 primer mixes used for DRB1\*16 SSP subtyping**

Primer Mix	Size of spec. PCR product <sup>1</sup>	Size of control band <sup>2</sup>	Amplified DRB1*16 alleles <sup>3</sup>	Other amplified DRB1 alleles <sup>4</sup>
<b>1</b>	260 bp	<b>515 bp</b>	*16:01:01-16:05:02, 16:07-16:14, 16:16-16:22, 16:24	*15:02:01-15:02:12, 15:08, 15:11, 15:14-15:15, 15:19, 15:26-15:27, 15:29-15:31, 15:34, 15:38-15:39, 15:44, 15:47, 15:58, 15:60, 15:63, 15:68, 15:78, 15:80N, 15:99, 15:101, 15:103-15:105
<b>2</b>	200 bp	<b>515 bp</b>	*16:02:01-16:02:03, 16:10-16:12, 16:14, 16:16-16:23	
<b>3<sup>6</sup></b>	200 bp	430 bp	*16:01:01-16:01:05, 16:03-16:04, 16:08-16:09:02, 16:13N, 16:15, 16:24	
<b>4</b>	215 bp	430 bp	*16:03	
<b>5</b>	220 bp	430 bp	*16:04, 16:18	*15:21
<b>6</b>	200 bp	430 bp	*16:05:01-16:05:02, 16:07	*15:10, 15:21
<b>7</b>	160 bp	<b>515 bp</b>	*16:07	
<b>8<sup>5</sup></b>	110 bp 175 bp	430 bp	*16:08 *16:14	
<b>9</b>	140 bp	430 bp	*16:09:01-16:10	*15:01:01:01-15:01:21, 15:01:23-15:02:09, 15:02:11-15:06:02, 15:08, 15:10, 15:12-15:33, 15:35-15:47, 15:49-15:58, 15:60-15:68, 15:70-15:87, 15:89-15:95, 15:97-15:112
<b>10<sup>5</sup></b>	115 bp	430 bp	*16:09:01-16:10	*11:01:03, 11:01:10-11:01:11, 11:04:07, 11:08:03, 11:19:02, 12:01:01, 12:01:03-12:02:03, 12:02:05-12:10, 12:12-12:15, 12:16:02-12:20, 12:23-12:37, 12:39-12:42, 12:44-12:48, 13:02:02, 13:77, 13:163, 13:181, 15:50N, 15:80N
<b>11</b>	170 bp 215 bp	430 bp	*16:21N *16:11	
<b>12</b>	215 bp	<b>515 bp</b>	*16:12 <sup>w</sup> , 16:17	
<b>13<sup>5</sup></b>	120 bp 165 bp	430 bp	*16:19 *16:13N, 16:14	
<b>14<sup>5</sup></b>	85 bp	430 bp	*16:16	*11:01:03, 11:01:10-11:01:11, 11:04:07, 11:08:03, 11:19:02, 12:04, <b>DRB5*01:13</b>
<b>15<sup>5</sup></b>	170 bp 80 bp	430 bp	*16:21N *16:15, 16:23	*01:23, 04:53, 04:99, 11:04:07,

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	12:01:01, 12:01:03-12:02:03, 12:02:05-12:06, 12:08-12:15, 12:17-12:21, 12:23-12:38, 12:41- 12:48, 13:77, 13:163, 13:181
<b>16<sup>7</sup></b>	<b>Negative Control</b>

<sup>1</sup>Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DRB\*16 SSP subtypings.

When the primers in a primer mix can give rise to HLA-specific PCR products of more than one length this is indicated if the size difference is more than 20 base pairs. Size differences of 20 base pairs or less are not given. For high resolution SSP kits, the alleles listed are specified according to amplicon length.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

<sup>2</sup>The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

<sup>3</sup>For several DRB1 alleles 1<sup>st</sup> and/or 3<sup>rd</sup> exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups.

<sup>4</sup>Due to the sharing of sequence motifs between DRB1 alleles, primer mixes 1, 5, 6, 9, 10, 14 and 15 will amplify other DRB1 alleles. In addition, DRB5 alleles will be amplified by primer mix 14.

<sup>5</sup>HLA-specific PCR products shorter than 125 base pairs have a lower intensity and are less sharp than longer PCR products.

<sup>6</sup>Primer mix 3 has a tendency to giving rise to primer oligomer formation.

<sup>7</sup>Primer mix 16 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by the control primer pairs matching the human growth hormone gene. HLA-specific PCR product sizes range from 75 to 200 base pairs and the PCR product generated by the HGH positive control primer pair is 430 base pairs.

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## PRIMER SPECIFICATION

Well No.	1	2	3	4	5	6	7	8	9	10	11	12
Length of spec. PCR product	260	200	200	215	220	200	160	110	140	115	170	215
Length of int. pos. control <sup>1</sup>	515	515	430	430	430	430	515	430	430	430	430	515
5'-primer(s) <sup>2</sup>	13(126) 5'-Agg 3'	13(126) 5'-Agg 3'	13(126) 5'-Agg 3'	13(126) 5'-Agg 3'	13(126) 5'-Agg 3'	13(126) 5'-Agg 3'	27(167) 5'-CCC 3'	13(126) 5'-Agg 3'	13(126) 5'-Agg 3'	47(227) 5'-gTT 3'	13(127) 5'-ggA 3'	13(126) 5'-AAG 3'
											29(173) 5'-Ag 3'	15(133) 5'-gTA 3'
3'-primer(s) <sup>3</sup>	86(344) 5'-CAC 3'	66(286) 5'-gAg 3'	66(286) 5'-gAA 3'	71(301) 5'-ggC 3'	73(307) 5'-CAg 3'	66(286) 5'-gAT 3'	66(286) 5'-gAT 3'	37(197) 5'-CgT 3'	47(227) 5'-ggA 3'	72(303) 5'-gCg 3'	72(303) 5'-gCg 3'	72(303) 5'-gCg 3'
			66(286) 5'-gAA 3'			66(286) 5'-gAT 3'	66(286) 5'-gAT 3'	57(258) 5'-gCT 3'				
Well No.	1	2	3	4	5	6	7	8	9	10	11	12

Well No.	13	14	15
Length of spec. PCR product	120	85	80
Length of int. pos. control <sup>1</sup>	430	430	430
5'-primer(s) <sup>2</sup>	13(126) 5'-Agg 3'	29(173) 5'-Ag 3'	72(303) 5'-CgC 3'
		58(261) 5'-gAg 3'	
3'-primer(s) <sup>3</sup>	39(203) 5'-AgT 3'	72(303) 5'-gCg 3'	86(344) 5'-CCA 3'
	51(241) 5'-CTA 3'		
	57(258) 5'-gCT 3'		
Well No.	13	14	15

<sup>1</sup>The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

<sup>2</sup>The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

<sup>3</sup>The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.



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CELL LINE VALIDATION SHEET														
DRB1*16 SSP subtyping kit <sup>2</sup>														
					Well									
					1	2	3	4	5	6	7	8	9	10
					11	12	13	14	15	16	17	18	19	20
					21	22	23	24	25	26	27	28	29	30
					31	32	33	34	35	36	37	38	39	40
					41	42	43	44	45	46	47	48	49	50
					51	52	53	54	55	56	57	58	59	60
					61	62	63	64	65	66	67	68	69	70
					71	72	73	74	75	76	77	78	79	80
					81	82	83	84	85	86	87	88	89	90
					91	92	93	94	95	96	97	98	99	100
					101	102	103	104	105	106	107	108	109	110
					111	112	113	114	115	116	117	118	119	120
					121	122	123	124	125	126	127	128	129	130
					131	132	133	134	135	136	137	138	139	140
					141	142	143	144	145	146	147	148	149	150
					151	152	153	154	155	156	157	158	159	160
					161	162	163	164	165	166	167	168	169	170
					171	172	173	174	175	176	177	178	179	180
					181	182	183	184	185	186	187	188	189	190
					191	192	193	194	195	196	197	198	199	200
					201	202	203	204	205	206	207	208	209	210
					211	212	213	214	215	216	217	218	219	220
					221	222	223	224	225	226	227	228	229	230
					231	232	233	234	235	236	237	238	239	240
					241	242	243	244	245	246	247	248	249	250
					251	252	253	254	255	256	257	258	259	260
					261	262	263	264	265	266	267	268	269	270
					271	272	273	274	275	276	277	278	279	280
					281	282	283	284	285	286	287	288	289	290
					291	292	293	294	295	296	297	298	299	300
					301	302	303	304	305	306	307	308	309	310
					311	312	313	314	315	316	317	318	319	320
					321	322	323	324	325	326	327	328	329	330
					331	332	333	334	335	336	337	338	339	340
					341	342	343	344	345	346	347	348	349	350
					351	352	353	354	355	356	357	358	359	360
					361	362	363	364	365	366	367	368	369	370
					371	372	373	374	375	376	377	378	379	380
					381	382	383	384	385	386	387	388	389	390
					391	392	393	394	395	396	397	398	399	400
					401	402	403	404	405	406	407	408	409	410
					411	412	413	414	415	416	417	418	419	420
					421	422	423	424	425	426	427	428	429	430
					431	432	433	434	435	436	437	438	439	440
					441	442	443	444	445	446	447	448	449	450
					451	452	453	454	455	456	457	458	459	460
					461	462	463	464	465	466	467	468	469	470
					471	472	473	474	475	476	477	478	479	480
					481	482	483	484	485	486	487	488	489	490
					491	492	493	494	495	496	497	498	499	500
					501	502	503	504	505	506	507	508	509	510
					511	512	513	514	515	516	517	518	519	520
					521	522	523	524	525	526	527	528	529	530
					531	532	533	534	535	536	537	538	539	540
					541	542	543	544	545	546	547	548	549	550
					551	552	553	554	555	556	557	558	559	560
					561	562	563	564	565	566	567	568	569	570
					571	572	573	574	575	576	577	578	579	580
					581	582	583	584	585	586	587	588	589	590
					591	592	593	594	595	596	597	598	599	600
					601	602	603	604	605	606	607	608	609	610
					611	612	613	614	615	616	617	618	619	620
					621	622	623	624	625	626	627	628	629	630
					631	632	633	634	635	636	637	638	639	640
					641	642	643	644	645	646	647	648	649	650
					651	652	653	654	655	656	657	658	659	660
					661	662	663	664	665	666	667	668	669	670
					671	672	673	674	675	676	677	678	679	680
					681	682	683	684	685	686	687	688	689	690
					691	692	693	694	695	696	697	698	699	700
					701	702	703	704	705	706	707	708	709	710
					711	712	713	714	715	716	717	718	719	720
					721	722	723	724	725	726	727	728	729	730
					731	732	733	734	735	736	737	738	739	740
					741	742	743	744	745	746	747	748	749	750
					751	752	753	754	755	756	757	758	759	760
					761	762	763	764	765	766	767	768	769	770
					771	772	773	774	775	776	777	778	779	780
					781	782	783	784	785	786	787	788	789	790
					791	792	793	794	795	796	797	798	799	800
					801	802	803	804	805	806	807	808	809	810
					811	812	813	814	815	816	817	818	819	820
					821	822	823	824	825	826	827	828	829	830
					831	832	833	834	835	836	837	838	839	840
					841	842	843	844	845	846	847	848	849	850
					851	852	853	854	855	856	857	858	859	860
					861	862	863	864	865	866	867	868	869	870
					871	872	873	874	875	876	877	878	879	880

101.126-12 – including *Taq* polymerase, IFU-01  
101.126-12u – without *Taq* polymerase, IFU-02

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“Instructions for Use” (IFU)

**Lot No.: 60X**

**Lot-specific information**

<sup>2</sup>The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solutions 4, 6 to 8, 11 to 14 were available. The specificities of the primers in primer solutions 6, 8 and 14 were tested by separately adding one additional 5'-primer, respectively one additional 3'-primer. In primer solutions 4 and 13 it was only possible to test the 5'-primers, the 3'-primers were not possible to test. In primer solution 7, 11 and 12 it was only possible to test the 3'-primers, the 5'-primers were not possible to test. In primer solution 14, one 5'-primer was not possible to test, and in primer solutions 3 and 8 one 3'-primer was not possible to test.

101.126-12 – including *Taq* polymerase, IFU-01

101.126-12u – without *Taq* polymerase, IFU-02

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101.126-12u – without *Taq* polymerase, IFU-02

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Lot No.: **60X**

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